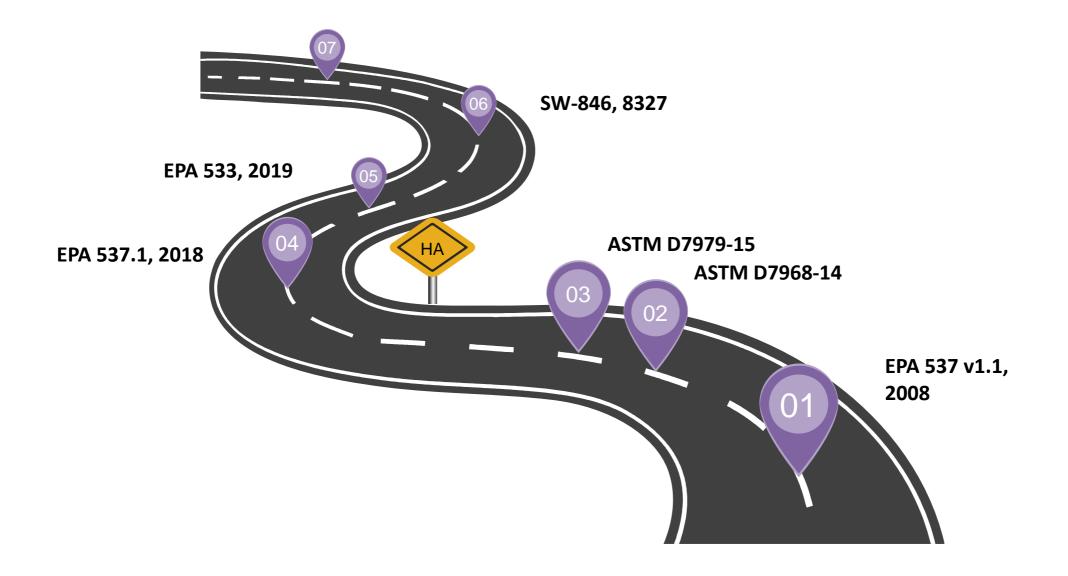


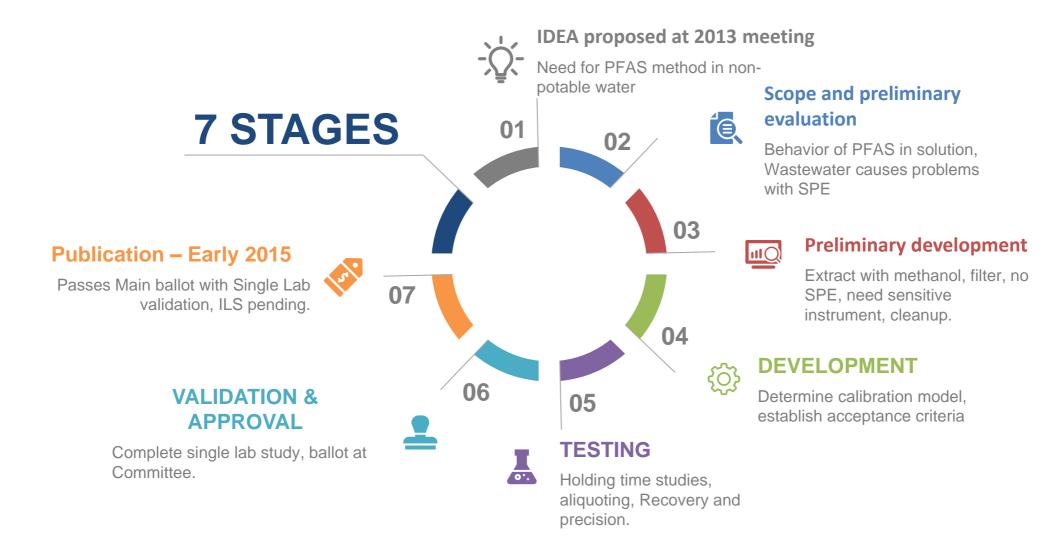
Analysis of Per and Polyfluorinated Alkyl Substances in non-Potable Water by co-solvation with methanol followed by tandem liquid chromatography mass spectrometry

> William Lipps Analytical and Measurement Division August 2020

A long winding road of PFAS methods, each for different purposes



ASTM D7979 first published in 2015 for analysis of non-potable water, including wastewater

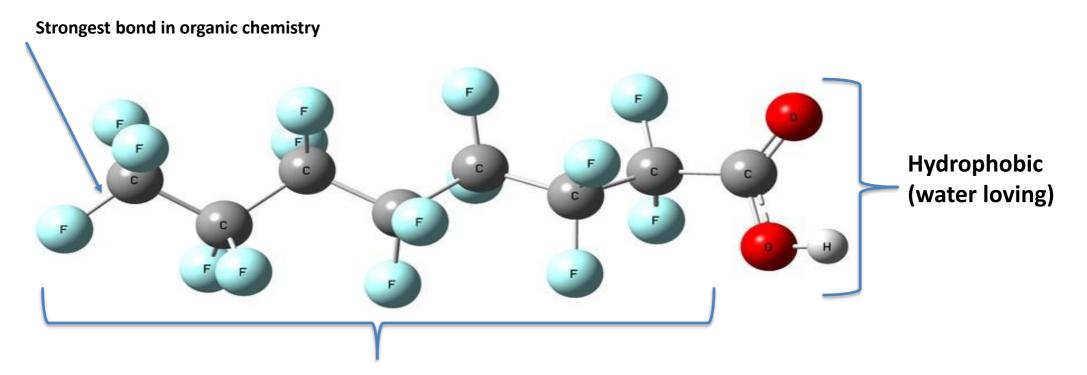


Multiple different facets are considered near simultaneously during method development



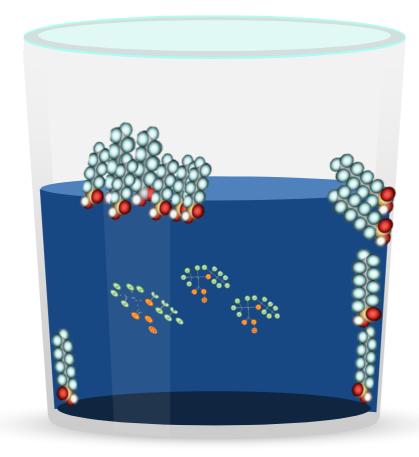
PowerPoint makes it linear

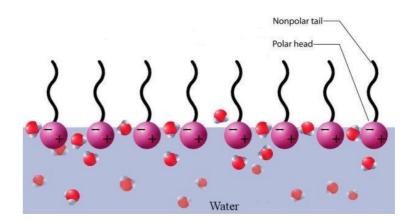
PFAS have unique chemical properties that must be considered



Hydrophobic (water repelling) , oleophobic (oil repelling), lipophobic (fat repelling)

PFAS compounds concentrate at interfaces



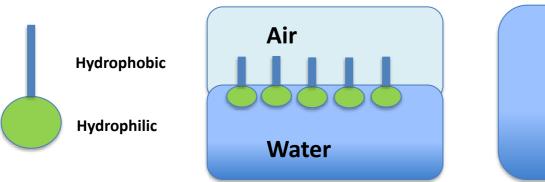


Bigger chains rise to the top faster

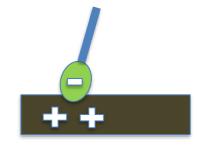
Smaller chains with more polar groups stay in solution longer

Polar heads – attach to surface

PFAS behaves like surfactants in solution



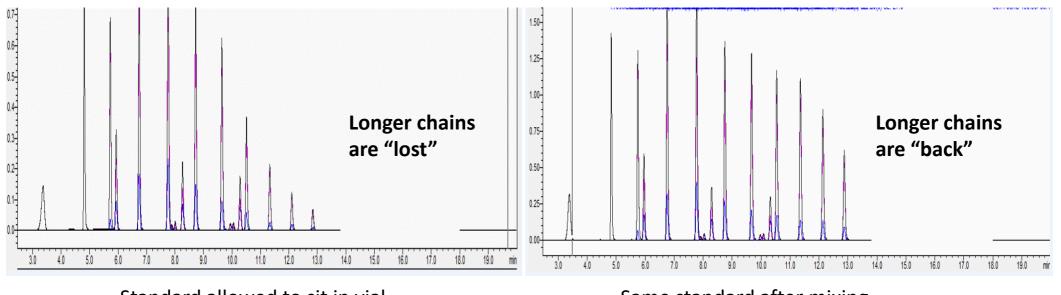
The C-F bond = weak Van der Waals and Hydrogen bonding



Critical Micelle Concentration(CMC) lowered by interaction with particles, organics, and soil. The negative charged head can interact with positive ions

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The loss of PFAS upon standing in a vial

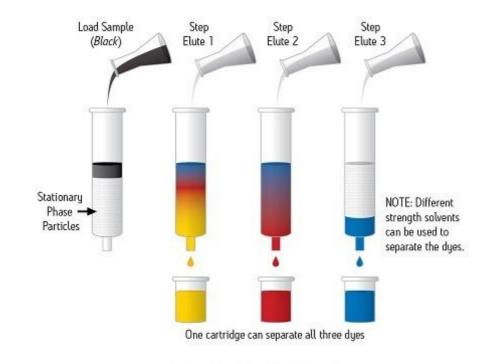


Standard allowed to sit in vial

Same standard after mixing

PFAS standards are "lost" upon sitting, but recovered by mixing, they "float". Potential for loss in subsampling

Existing Method 537 v1.1 used SPE - evaluation of Solid Phase Extraction



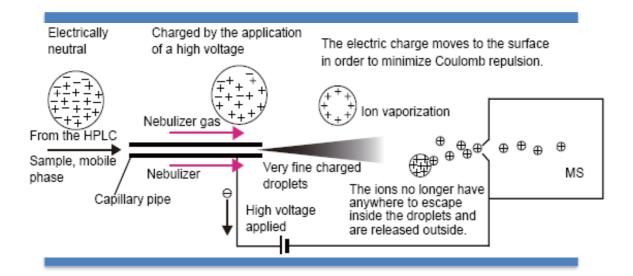
https://www.waters.com/waters/en_US/Solid-Phase-Extraction-SPE-Guide/nav.htm?cid=134721476&locale=en_US

Benefits of SPE

- Compound Purification removal of complex matrix
 - Reduce ion suppression or enhancement – MS
- **X** 3. Fractionate compounds by class
- A. Transfer sample from aqueous to organic phase (GC methods)
- **?** 5. Enrichment of analyte concentration

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Suppression or enhancement at the interface, investigation of cause and potential remedies



Matrix effects \rightarrow coeluting compounds alter ionization energy at the interface

Improved separation best technique to overcome

Interferences usually elute early – divertor valve Lower flow rates Enhancement = increase in signal Suppression = decrease in signal

Phospholipids main cause

Higher % solvent, lowers surface tension and boiling point resulting in more efficient de-solvation.

Weakly acidic (ammonium acetate) volatile solvent. Acetonitrile better then methanol

With correct mobile phase, column and gradient, interferences are minimized for most wastewater samples

Evaluation of Solid Phase Extraction for wastewater and highly polluted samples

SW-846 3535 Solid Phase Extraction – Not appropriate for samples with TSS > 1%

- 1. Particulates may clog SPE media
- 2. For PFAS must wash out entire container
- 3. Preliminary filtration= two extractions per sample

Filtration **prior to** extraction = analyte loss

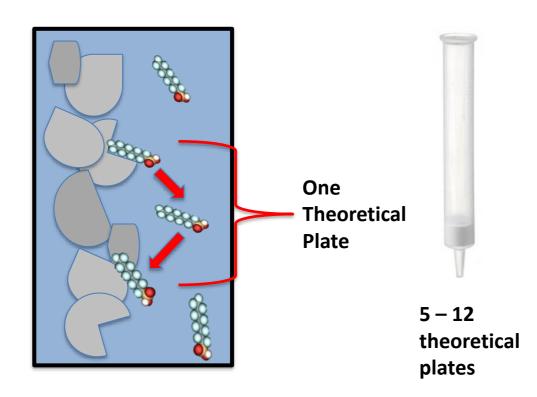
You want retention of targets and not interferences, but:

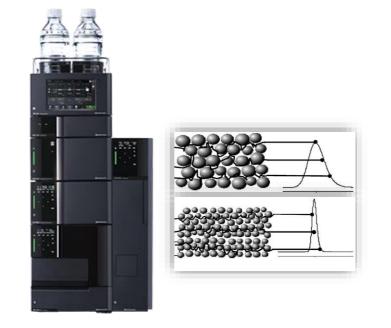
- 1. Interferences may also be retained and eluted with analytes
- 2. Depending on media, some analyte may not be retained
- 3. Depending on media, some analytes may not be recovered

Goal is to have one method for WW capable of extraction and analysis in a single, not two or more extractions.

Preliminary filtration not an option, and SPE requires more than one media to quantitatively recover all target PFAS (C4 – C14)

A LC Column can be used as, and is essentially, SPE

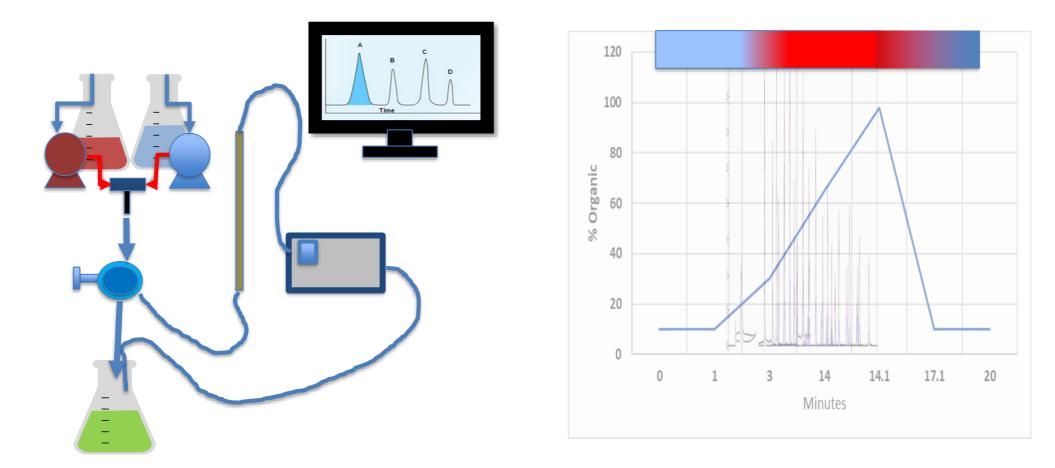




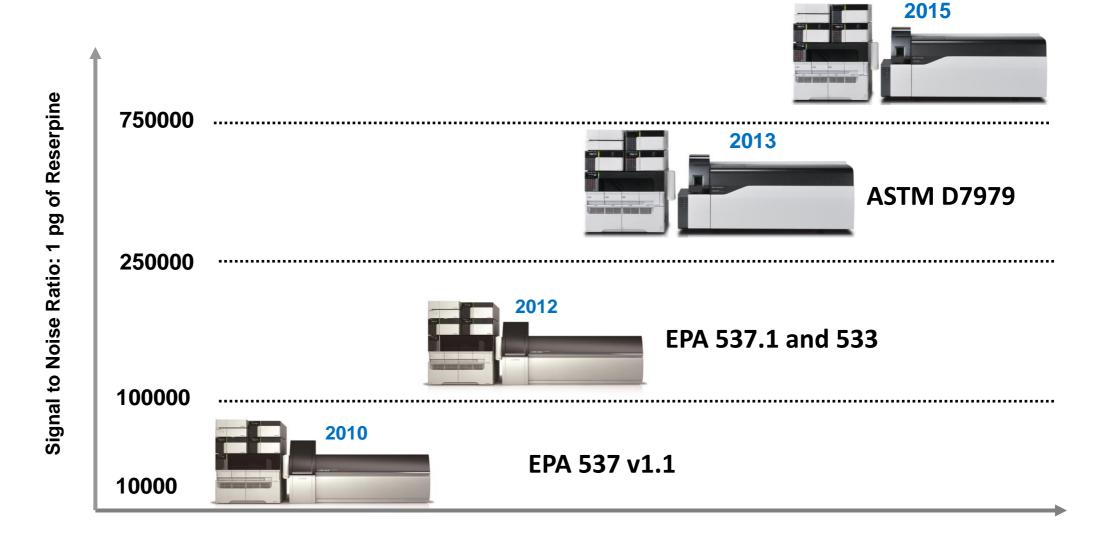
10,000- 20,000 theoretical plates

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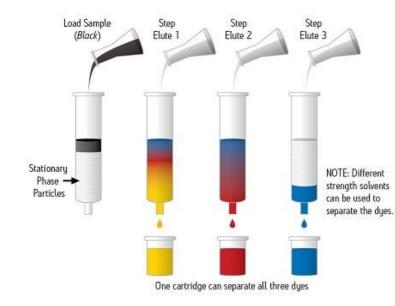
Correctly applied, the HPLC column serves as SPE



Enough sensitivity for 10 - 30 microliter injections, no preconcentration needed



Evaluation of Solid Phase Extraction, revisited



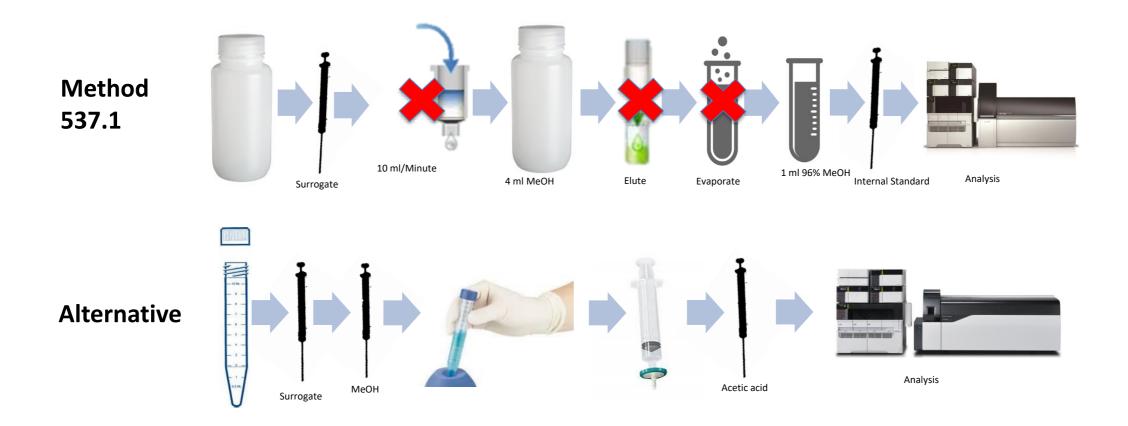
https://www.waters.com/waters/en_US/Solid-Phase-Extraction-SPE-Guide/nav.htm?cid=134721476&locale=en_US

Benefits of SPE

- Compound Purification removal of complex matrix
 - 2. Reduce ion suppression or enhancement – MS
- **X** 3. Fractionate compounds by class
- X 4. Transfer sample from aqueous to organic phase (GC methods)
 - 5. Enrichment of analyte concentration

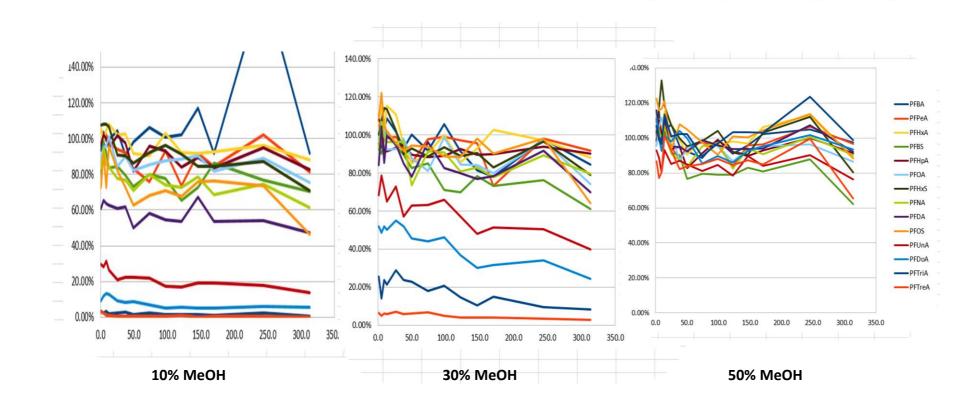
Potential interferences and need for analyte enrichment can be handled using modern instrumentation

Devise an extraction, verify if it works, if not change it



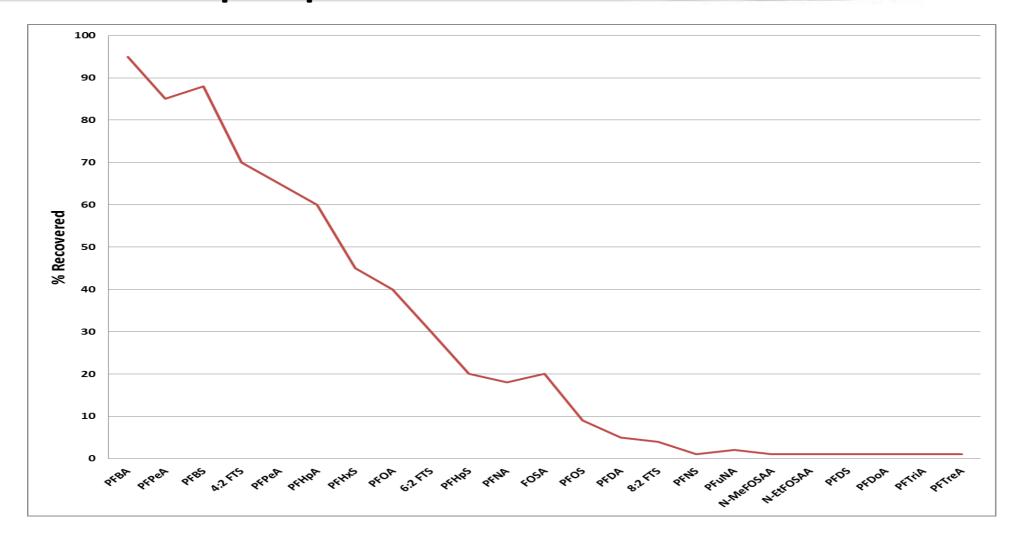
Consistent with ASTM sustainability and technology innovation goals Extraction is "like" making standards

Solubility and stability of analytes in methanol



Study demonstrates sample should be 50:50 Methanol and water for maximum solubility and minimum dilution. All standards and samples prepared in same way.

Activated carbon significantly removed analyte, so no clean-up step in the method.



Guidelines for determination of the appropriate calibration model, based on the single lab study

Recovery of mid range Spikes (all evaluated matrices)	RSD	Calibration Model
70 – 130 %	≤ 20 %	External Standard
< 70 %	≤ 20 %	External Standard ¹
> 130 %		Reject the analyte or modify the extraction
70 – 130 %	> 20%	Use internal standard calibration, or isotope dilution (if isotopes are available)

¹ External Standard Calibration is appropriate, however, consider rejecting the analyte or modifying the extraction

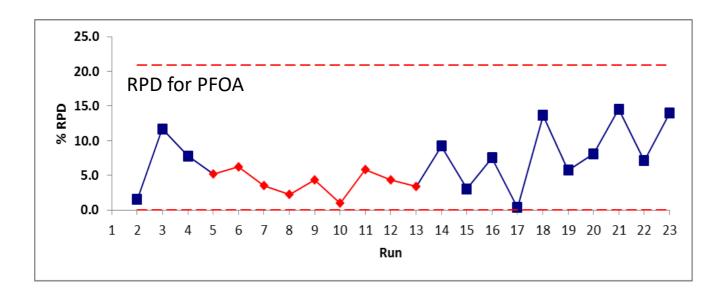
1 SHIMADZU

6 matrices met condition 1, therefore, external standard calibration was accepted

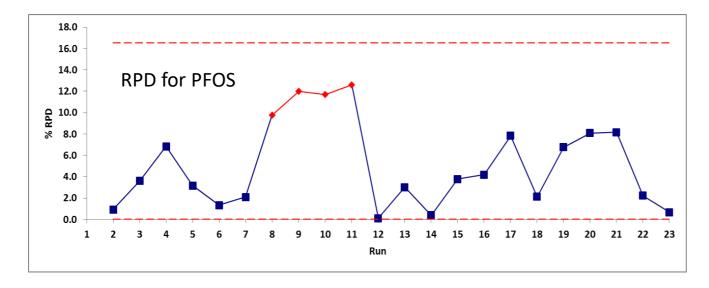
	Reagent Water			River water I	I	Sewage Plan	t IV Effluent	Sewage Plant IV Influent	Lake Water		Ground Water		Average across Matrices		
Unlabled Native Analyte	Concentration (ng/L)	% Recove	y % RSD	% Recovery	% RSD	% Recovery	% RSD	% Recovery	% RSD	% Recovery	% RSD	% Recovery	% RSD	% Recovery	% RSD
PFTreA		160 91	.9 8.	5 100	4.04	82.7	4.75	81.7	2.44			93.6	2.31	. 91.	5 4.02
PFTriA		160 91	.7 4.4	4 104	2.57	88.5	3.79	88.9	2.9	99.6	2.32	2 98	3.16	95.:	1 3.19
PFDoA		160 92	.2	4 100	1.94	90.3	3.36	91.1	2.48	98.8	1.93	97.3	2.12	95.0	2.64
PFUnA		160 92	.9 2.	5 97.1	2.16	89.8	2.84	91.3	4.54	94.4					
PFDA		160 93	.7 1.	5 96.8	2.81	. 89.4	4	95.8	3.39	96.6	2.61	93.3	3.1	. 94.3	3 2.90
PFNA		160 94	.4 2.3	2 97.6	3.17	90.7	3.46	94.3	4.05	96.8	1.73	96.6	5 1.25	95.3	1 2.64
PFOA		160 91	.1 2.3	3 90.6	3.52	92.2	3.02	101	3.72	92.5	3.25	5 90.8	2.62	93.0	3.07
PFHpA		160 65	.2 7.4	4 95.5	2.16	85.8	3.89	89.8	4.35	91.8	2.43	92.5	1.69	86.8	3.65
PFHxA		160	58 1.9	9 92.9	2.49	87	2.31	96.5	4.5	91.5	4.15	96.5	2.24	87.:	1 2.93
PFPeA		800 88	.1 1.4	4 95.2	1.09	82.6	3.28	81	3.45	84.1	7.45	94.4	3.27	87.0	5 3.32
PFBA		800 70	.4 1.	7 68.3	4.82	47.3	10.6	47.3	8.46	60.3	17.3	65.8	5.83	59.9	8.12
PFBS		160	99 17.9	9 93.4	2.74	84.7	3.67	90.5	5.05	87.3	4.15	98.9	1.65	92.3	3 5.86
PFHxS		160 92	.1 3.1	1 103	1.97	88.8	3.33	92.1	3.3	100	2.21	100	1.42	96.0	2.56
PFOS		160 94	.3 3.	3 102	2.79	91.7	2.56	95.1	4.03	103	3.22	2 102	. 1.74	. 98.0	3.02
PFDS		160 101	.4 4.2	4 109	1.3	92.7	2.42	90.3	2.48	107	1.66	5 103	3.1	. 10:	1 2.53
PFNS		160 99	.8 3.2	1 106	2.33	90.8	2.22		2.47	106	1.55	5 103	1.22	. 99.	5 2.17
PFHpS		160 98	.8 4.5	5 104	1.43	90.8	2.42	90.8	2.78	103	2.66	5 102	2.49	98.3	2 2.72
PFPeS		160 93			1.15		2.72		3.35						
FOSA		160 98	.3 2.5	4 93.6	1.42	92.7	2.3	80.4	1.63	94.9	1.52	92.3	1.43	92.0	1.81
4:2 FTS		160 99	.5 4.3	2 97.2	1.83	86.8	3.44	96.4	4.42	95.7	3.04	4 95	2.42	95.2	1 3.25
6:2 FTS		160 105	.1 13.:	1 102	4.4	89.8	2.76	91.5	4.01	102	3.56	5 97.3	3.9	98.0	5.29
8:2 FTS		160 111	.7 6.4	4 108	1.02	97.2	4.17	98.4	4.08	108	<u>د</u>	104	2.95	5 10	5 3.78
N-EtFOSAA		160 103	.4 4.8	3 106	3.83	95.8	3.1	96.2	3.71			104	2.91	. 102	2 3.73
N-MeFOSAA		160 101	.3 3.6	5 98.6	1.66	84	3.28	98	2.28	99.1	2.98	97.4	2.85	96.4	1 2.78
	Reagent Water				Sewage Plant IV Effluent Sewage Plant IV Influent				Ground Water		Average across Matrices				
Labeled Surrogate		% Recove	y % RSD	% Recovery	% RSD	% Recovery	% RSD	% Recovery	% RSD	% Recovery	% RSD	% Recovery	% RSD	% Recovery	% RSD
MPFBA		160 91	.1 2.1	3 72.9	3.93		7.25	45.7	7.22	62.8	15.6	5 70.1	4.24	65.	5 6.76
MPFHxA		160 98	.2 1.4	4 95.8	1.56	90.4	2.17	94	4.38	93.8	4.13	95.7	1.82	94.	7 2.58
MPFHxS		160 97	.5 2.8	8 103	1.91	. 93.3	2.25	94.3	3.67	102	3.1	102	2.33	98.	7 2.68
MPFOA		160 100	.2 1.3		1.7	92	2.44	94.1	4.01			96.1			
MPFNA		160 98	.9 2.	3 96.9	2.14	90.6	3.26	93.7	2.88	96.9	2.78	96.1	2.06	95.	5 2.57
MPFOS		160 97	.2 1.:	1 108	1.96	90.9	2.94	91.2	3.19	107	2.89	106	2.33	100	2.40
MPFDA		160 98	.1 1.	5 98.2	2.11	90.9	2.94	95.6	2.67	97.7	3.48	97	2	96.3	3 2.47
MPFUnA		160 97	.8 0.9	9 99	1.5	91	2.52	92.1	3.76	97.9	2.82	97.5	5 1.9	95.9	2.23
MPFDoA		160 96			1.38				2.27						
M 4:2 FTS		160 101	.7 4.9	2 99.8	2.62	84.6	2.64	91.6	5.59	97.8	3.71	98.7	1.28		
M 6:2 FTS		160 108			4.22		3.56		3.95						
M 8:2 FTS		160 107			3.65		5.23		4.55						
MNEtFOSAA			11 5.		2.04				3.94						
M NMeFOSAA		160 103			1.83		2.78		2.92						

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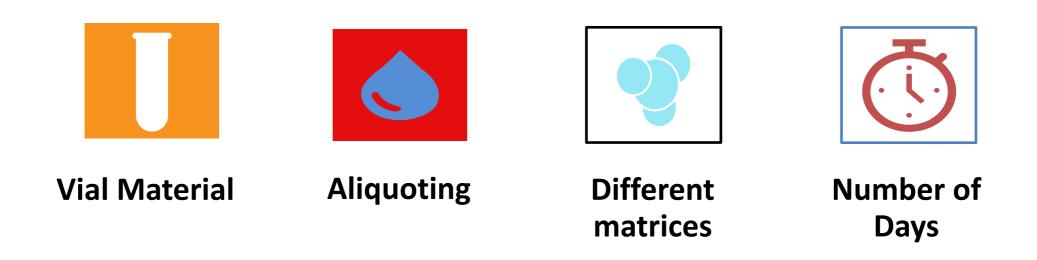
Evaluation of 5 ml samples size, precision in collocated samples, over one month



RPD of 5 ml samples of all matrices.

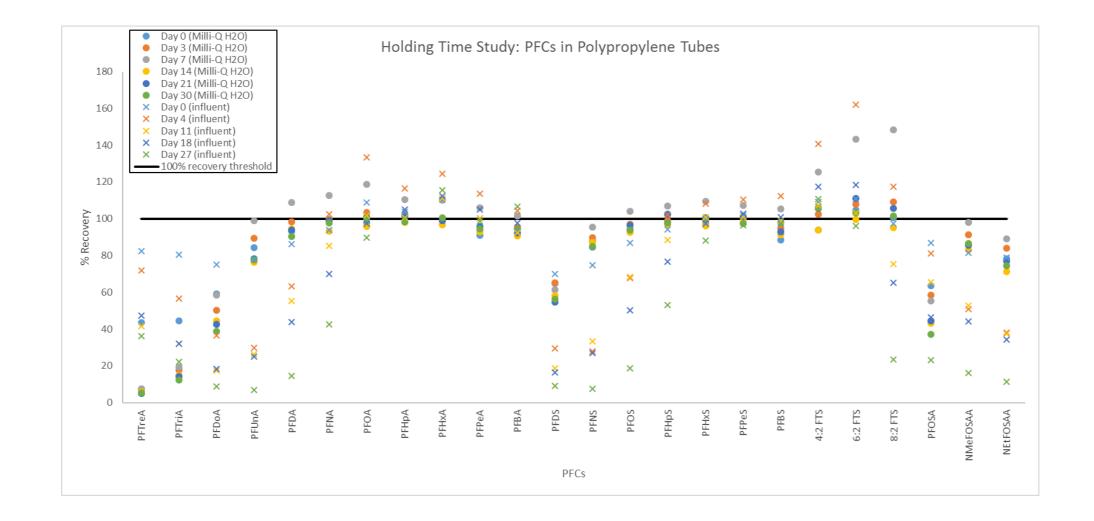


Holding time, vial material, and "aliquoting studies



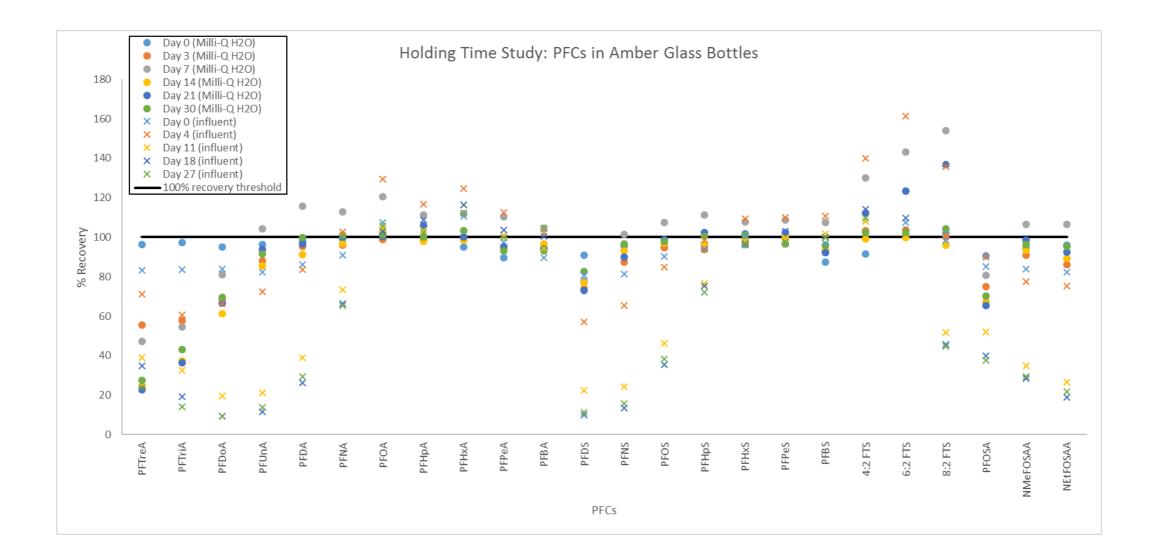
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Aliquoting in Polypropylene tubes



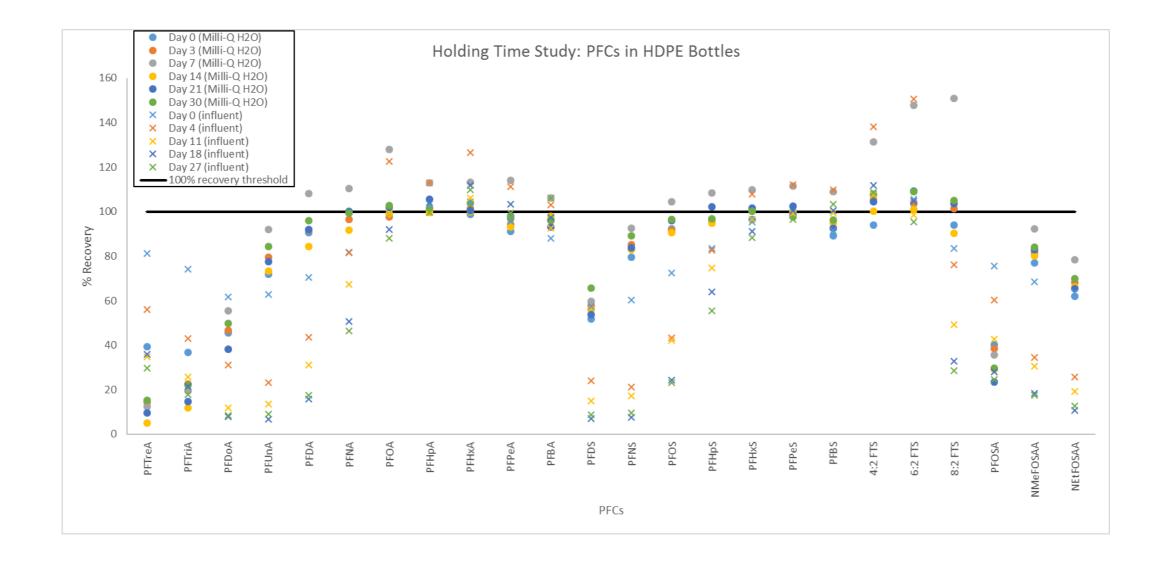


Aliquoting in amber glass bottles

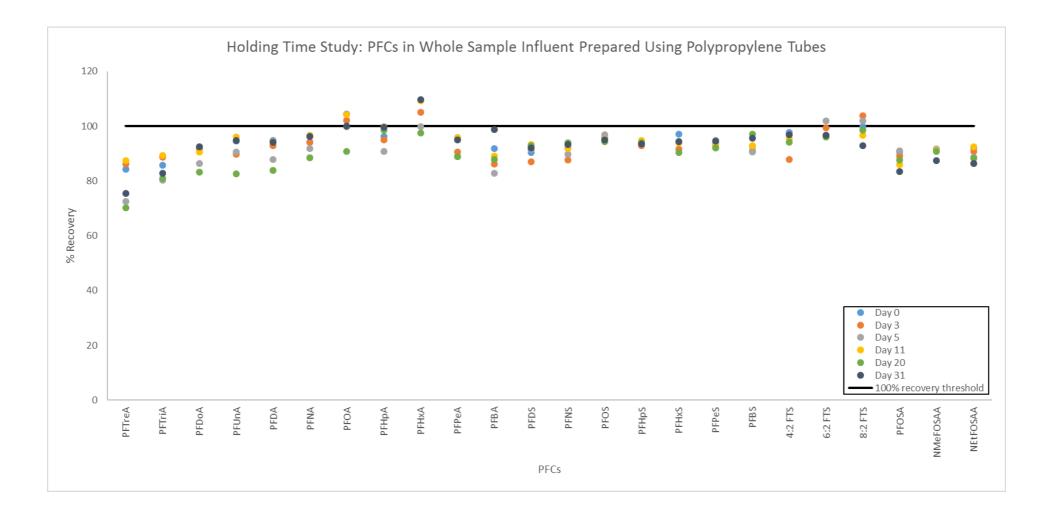




Aliquoting in HDPE bottles

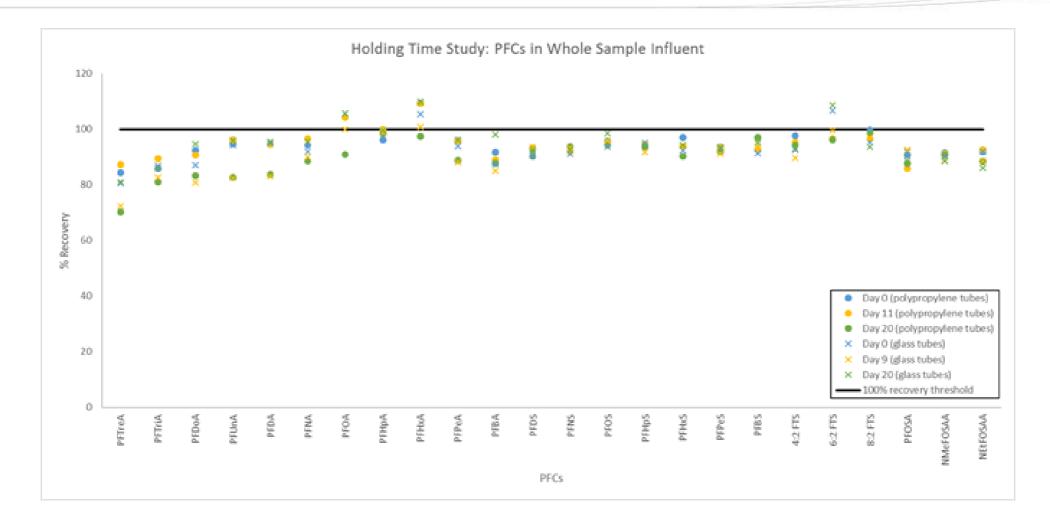


Whole sample "extracted" and analyzed from one Polypropylene (PP) tube, wastewater influent



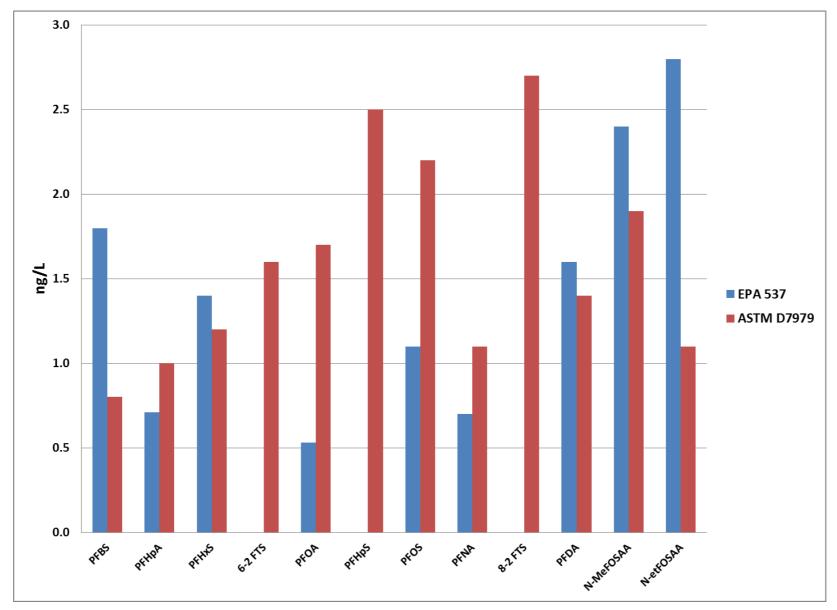
Acceptable recovery for all over 31 days

Whole sample "extracted" and analyzed from one glass tube, wastewater influent

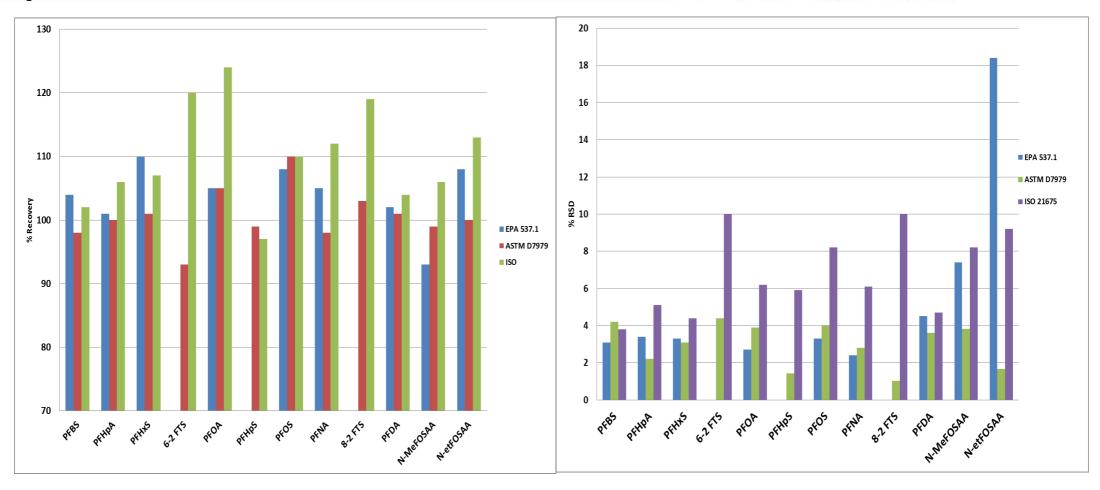


Acceptable recovery for all over 31 days, for safety D7979 uses PP

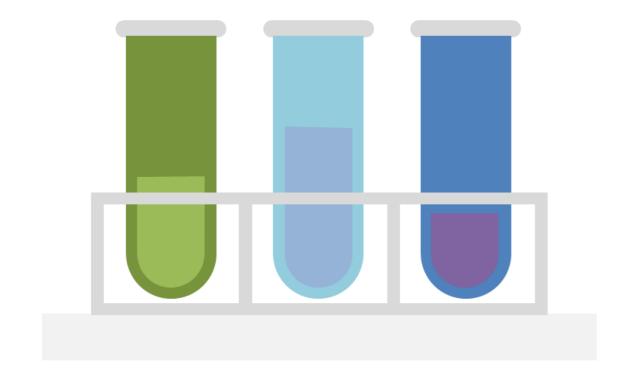
Comparison of D7979 detection limits with Method 537



Comparison of ASTM D7979 recovery and precision with EPA and ISO methods



Conclusion = D7979 validated for wastewater with comparable results to other PFAS methods





Validated in WW, with easy, "green" extraction



No SPE needed, filter after methanol added so high TSS not a factor



Detection limits, recovery, and precision comparable to other methods



Any Questions?

Contact Information

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